

Distance sampling: Methods and applications

Lure point transect case study: Scottish crossbills

Contents

1	The trials	1
2	The survey	3
3	Measure of precision	4
	References	5

The Scottish crossbill is Britain's only endemic bird species. A lure point transect survey was used to estimate its population size (Summers and Buckland 2011). In this case study, we work through the analyses reported in Section 10.2.1 of the book. We do not attempt to recreate those analyses exactly; instead, we simplify the analyses, while retaining the essential elements to allow the approach to be adapted to other studies.

1 The trials

The trials data are in file `lure_trials.csv`. We first read the data:

```
xbill <- read.table("lure_trials.csv", header = T, sep = ",")
attach(xbill)
```

We next specify a distance (in metres) beyond which it is reasonable to assume that no crossbill will respond to the lure:

```
w <- 850
```

There are several covariates in the data that might affect probability that a bird responds to the lure: `day` (days from 1st January); `time` (hour of the day); `habitat` (1 = plantation, 2 = native pinewood); `behavcode` (1 = perching and feeding, 2 = giving excitement calls, 3 = singing); `numbirds` (flock size); and `dist` (distance of birds from the point when the lure was played). We can fit all these covariates as follows.

```
habitat <- factor(habitat)
behavcode <- factor(behavcode)
model1 <- glm(response ~ dist + numbirds + day + time + habitat + behavcode,
             family = binomial)
summary(model1)
```

Call:

```
glm(formula = response ~ dist + numbirds + day + time + habitat +
     behavcode, family = binomial)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.9526	-0.8708	0.5563	0.8168	2.0009

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	2.641601	1.301148	2.030	0.0423 *

```

dist          -0.010086   0.002323  -4.342 1.41e-05 ***
numbirds      0.076882   0.081172   0.947  0.3436
day           -0.008597   0.007405  -1.161  0.2456
time          -0.020251   0.113258  -0.179  0.8581
habitat2      -0.263713   0.574532  -0.459  0.6462
behavcode2    0.070903   0.495256   0.143  0.8862
behavcode3    0.962426   0.614541   1.566  0.1173
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

(Dispersion parameter for binomial family taken to be 1)

```

Null deviance: 219.23 on 166 degrees of freedom
Residual deviance: 169.10 on 159 degrees of freedom
(8 observations deleted due to missingness)
AIC: 185.1

```

Number of Fisher Scoring iterations: 5

```
anova(model1)
```

Analysis of Deviance Table

Model: binomial, link: logit

Response: response

Terms added sequentially (first to last)

	Df	Deviance	Resid. Df	Resid. Dev
NULL			166	219.23
dist	1	44.961	165	174.27
numbirds	1	1.514	164	172.76
day	1	0.825	163	171.93
time	1	0.115	162	171.82
habitat	1	0.022	161	171.80
behavcode	2	2.694	159	169.10

We see that only `dist` has a coefficient that is significantly different from zero. Experiment with dropping non-significant terms. Using a backwards stepping procedure, you should find that other covariates remain non-significant, so we can use the simple model:

```

model2 <- glm(response ~ dist, family = binomial)
summary(model2)

```

Call:

```
glm(formula = response ~ dist, family = binomial)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.9521	-0.8066	0.5957	0.7618	1.9355

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	2.16370	0.33944	6.374	1.84e-10 ***
dist	-0.01049	0.00210	-4.993	5.94e-07 ***

```
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

(Dispersion parameter for binomial family taken to be 1)

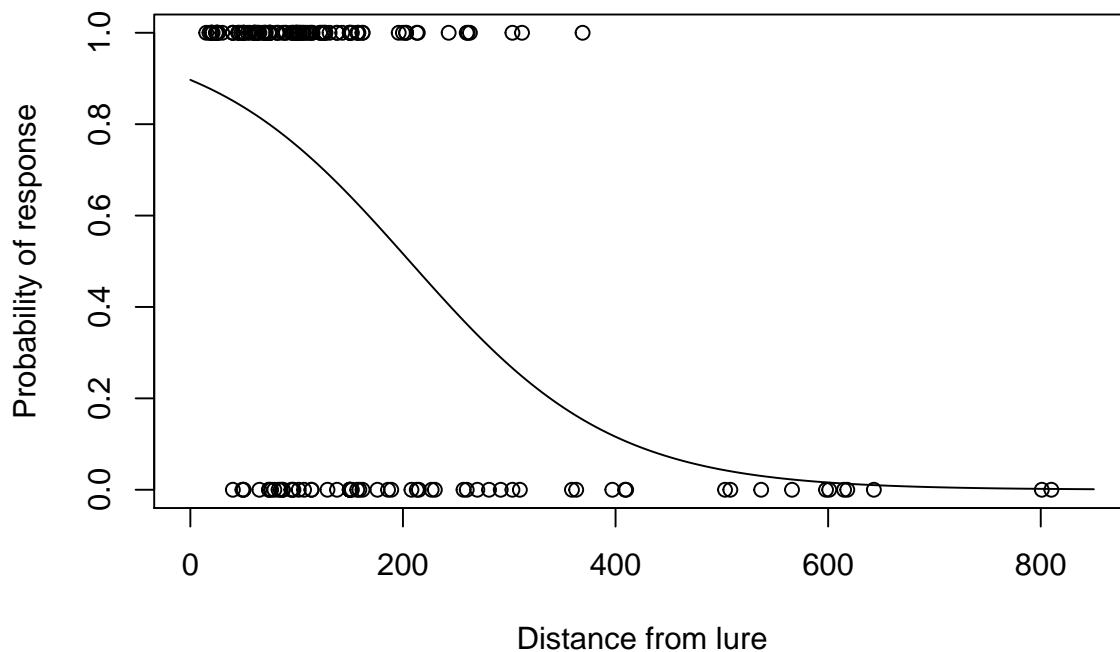
```
Null deviance: 227.52 on 174 degrees of freedom
Residual deviance: 179.44 on 173 degrees of freedom
AIC: 183.44
```

Number of Fisher Scoring iterations: 5

(Degrees of freedom change a little as some covariates have a few missing values.)

We can now plot the fitted detection function model:

```
fits <- fitted(model2)
plot(dist, response, xlim = c(0, w), xlab = "Distance from lure", ylab = "Probability of response")
dist2 <- 0:w
phat <- predict.glm(model2, newdata = data.frame(dist = dist2), type = "response")
lines(dist2, phat)
```



2 The survey

We need to specify the function $f(r) = \pi(r)/\pi(w)$, which represents the probability density function of distances of animals from the point. If we extend the study area out a distance w around the entire boundary, thus creating a bufferzone, and place our survey points at random throughout the area + bufferzone (or randomly place a systematic grid of points over the extended study area), we can assume that this density is triangular:

```
pi_r <- dist2/sum(dist2)
```

If the study area is fragmented, it may not be cost-effective to sample into the bufferzone. If points are restricted to within the boundaries of the study area, and the species of interest does not occur beyond the study area because habitat is unsuitable, we need to adjust $/pi(r)$ so that it reflects the amount of available habitat (averaged over points) with increasing distance from the point (Buckland et al. 2006). (Summers and Buckland 2011) estimated this availability function, but for the purposes of this case study, we use the above triangular distribution.

```
# Pa is prob of response, integrating r from 0 to w, assuming that phat is a
# function of distance alone (no z's in model)
```

```
(Pa <- sum(phat * pi_r))
```

```
[1] 0.09709381
```

Thus we estimate that just under 10% of birds within 850m of a point are detected. We now read in the data from the main survey. Note that detection distances are unknown in the main survey; instead, we have used the trials data to estimate the detection function, and hence the proportion of birds within 850m that are detected.

```
detections <- read.table("mainsurveydetections.csv", sep = ",", header = T)
attach(detections)
```

We now calculate the number of points k_{main} in the main survey, and the total area a within 850m of a point, converting from m^2 to km^2 . (Note that, if we do not include unsuitable habitat as described above, this total area should also exclude that habitat.)

```
kmain <- length(point)
a <- kmain * pi * (w/1000)^2
A <- 3505.8 # size in km2 of study area
```

We can now estimate the size of the population as

```
(Nscot <- sum(nscottish) * A/(Pa * a))
```

```
[1] 10007.67
```

Thus we estimate that there are around 10,000 birds. (This does not agree with the estimate of 13,600 in (Summers and Buckland 2011), because we have opted to keep the case study simple, and have omitted corrections for unsuitable habitat on surveyed plots, for juveniles, for unidentified crossbills, for incubating or brooding females, and for flying birds. Correcting only for unsuitable habitat on surveyed plots increases the abundance estimate to 12,900 birds.)

3 Measure of precision

We can calculate a standard error for our estimate and a 95% confidence interval for true abundance by bootstrapping both trials and points:

```
index <- 1:kmain
nboot <- 999
bNscot <- vector(length = nboot)
m <- length(dist)
tindex <- 1:m
bdist <- vector(length = m)
bresponse <- vector(length = m)
```

```
for (i in 1:nboot) {  
  # bootstrap trials  
  btindex <- sample(tindex, m, replace = TRUE)  
  for (l in tindex) {  
    bdist[l] <- dist[btindex[l]]  
    bresponse[l] <- response[btindex[l]]  
  }  
  bmodel <- glm(bresponse ~ bdist, family = binomial)  
  bphat <- predict.glm(bmodel, newdata = data.frame(bdist = dist2), type = "response")  
  bPa <- sum(bphat * pi_r)  
  # bootstrap points  
  rindex <- sample(index, kmain, replace = TRUE)  
  bNscot[i] <- sum(nscottish[rindex]) * A/(bPa * a)  
}  
# calculate bootstrap standard error  
seNscot <- sd(bNscot)  
# calculate bootstrap percentile confidence limits  
bNscot <- sort(bNscot)  
alpha.over.2 <- 0.025  
loNscot <- bNscot[round(alpha.over.2 * (nboot + 1))]  
hiNscot <- bNscot[round((1 - alpha.over.2) * (nboot + 1))]  
(c(Nscot, seNscot, loNscot, hiNscot))
```

```
[1] 10007.672 2855.774 5693.588 16793.749
```

Thus we estimate the population size to be 10008 birds with standard error 2856 and 95% confidence interval (5694, 16794).

References

Buckland, S. T., R. W. Summers, D. L. Borchers, and L. Thomas. 2006. Point transect sampling with traps or lures. *Journal of Applied Ecology* 43:377–384.

Summers, R. W., and S. T. Buckland. 2011. A first survey of the global population size and distribution of the Scottish Crossbill *loxia scotica*. *Bird Conservation International* 21:186–198.

This document describes a case study from

Distance Sampling: Methods and Applications
published by Springer

See [Case studies website](#)

Also see [Distance sampling website](#)
